• 药物分析与检验 •

Measurement of cyclosporine A in aqueous humor of rabbit eyes by HPLC

LIU Ai-ming¹, YANG Ju-lin², Lü Zhi-hua² (1. Shandong Eye Hospital and Institute, Qingdao 266071, China; 2. Chinese Ocean Universety, Qingdao 266003, China)

ABSTRACT: OBJECTIVE To develop a simple and reproducible high-performance liquid chromatography (HPLC) method for determination of cyclosporine A (CsA, also known as cyclosporin A) in aqueous hum or of rabbit eyes using CsD as internal standard. METHODS The pretreatment involved one step of liquid-liquid extraction with ether from aqueous hum or samples. Chromatography was carried out using an ODS column which was controlled at 75°C under isocratic elution with the mobile phase of acetonitrile-deionized water (89:11, v/v) at the flow rate of 0.1 mL• m in and the detector set at 200 nm. RESULTS The mean absolute recovery of CsA from aqueous hum or was 86% and the mean relative recovery was 100.7%. The linearity was assessed in the range of 25 ~500 ng mL with a correlation coefficient of 0.9986. The limit of quantification and detection of the present method were 25 and 10 ng• mL new materials. This method has been used to analyze large numbers of aqueous hum or samples of rabbit eyes for ophthalm ic research and proved to be rapid with good accuracy and precision.

KEY WORDS: cyclosporine A; HPLC; aqueous hum or of rabbit eyes

反相高效液相色谱法测定兔眼房水环孢素 A

刘爱明¹,杨菊林²,吕志华²(1.山东省眼科研究所,山东 青岛 266071; 2.中国海洋大学,山东 青岛 266003)

摘要:目的 以环孢素 D为内标物,建立 RP-HPLC法测定兔眼房水环孢素 A(CsA)药物浓度。方法 兔眼房水标本经乙醚单步提取挥干后,色谱过程使用 ODS柱,乙腈:水(89:11)为流动相,流速:0.1 mL• min⁻¹,75℃柱温下进行色谱分析,紫外检测波长为 200 nm。结果 方法的绝对回收率平均为 86%,相对回收率平均为 100.7%, CsA兔眼房水浓度在 25~500 ng• mL⁻¹范围内呈良好的线性关系,r=0.9986,最低定量限与最低检测限分别为 25 ng• mL⁻¹,10 ng• mL⁻¹。结论 眼科药物研究过程中大量房水标本检测实践表明,本方法过程简便、快速、结果准确、精密度符合方法学要求。

关键词:环孢素 A;反相高效液相色谱法;兔眼房水

中图分类号: R917.795.01 文献标识码: B 文章编号:1007-7693(2005)04-0301-03

1 Introduction

Cyclosporine is a cyclic undecapeptide consisting of 11 am in no acids. It is an important immunosuppressive drug used commonly to reduce tissue rejection after organ transplantation. Topical preparation of CsA drug delivery system vehicled by biodegradable PLGA or ophthalm ic solution can be used to prevent rejection of penetrating keratoplasty (PKP)^[1,2]. Methodological approaches to measuring CsA include high-performance liquid chromatography (HPLC) and a variety of immunoassay-based methods. Although immunoassay-based methods are simple to use, the cross-reactivity of the antibody with the metabolites leads to overestimation of the parent CsA concentrations in samples by these methods. The great different amount of protein in blood and aqueous hum or make them not appropriate for determination of CsA in aqueous hum or because of the different absorp-

tion[3~5]

During the past two decades, many attempts have been made to improve HPLC method for CsA analysis in blood, resulting in numerous published methods most of which involve the use of solid-phase extraction (SPE) procedure or liquid liquid extraction followed by reconstitution with n-hexane using Cyclosporin D (CsD) as internal standard, which is time-consuming ^{6,71}. What's more, the described chromatographic operation can only offer detection limit of 50 ng• mL⁻¹ from 1 to 2mL matrix which can not be followed to determine the small-volumed specimen with low CsA concentration such as aqueous humor with volume of 0.2 mL.

The present study aims to modify the reported method for CsA assay in aqueous humor. In this paper, we report a simple and reproducible HPLC method using simple pretreatment proce-

作者简介:刘爱明, 29岁,硕士,主管药师。 Tel: 0532 - 85876380-163. E-mail: aim ing-Liu0105@ sina. com

dure and demonstrate applicability of the method.

2 Materials and method

2.1 Reagents

All chemicals were reagent or HPLC grade. Acetonitrile, methanol, were obtained from Merck, Germany. Both CsA and CsD standard products were provided by National Institute for Control of Pharmaceutical and Biological Products. Deionized water was obtained by using USF-ELGA purification system.

2.2 Instrumentation

Analysis was perfomed using a Agilent 1100 HPLC system (Agilent, USA) consisting of a on-line degasser, a quaternary pump, a autoinjector, a column oven, a VWD detector and a chem station. A stablebond C_{18} reversed-phase column (80 Å, 5 μ m, 2.1 mm i. d., dpl 50 mm ZOBARX, Agilent, USA) was used for chromatographic separation. The detector was set to 200 nm. The mobile phase comprised of aceton itrile-deionized water (89:11). Analyses were run at flow rate of 0.1 mL · m in -1 at 75 °C for 10 m in. The samples were quantified using peak height ratio of CsA and CsD.

2.3 Standard solutions

The stock standard solutions of CsA and CsD (1 mg • mL $^{-1}$) were prepared by dissolving the powder in methanol respectively. For constructing calibration curves and for assay validation covering the range, both the working standard solutions of 2.5 μ g • mL $^{-1}$ for CsA and 25 μ g • mL $^{-1}$ for CsD were obtained by diluting stock solutions.

2.4 Sample preparation

To a 200 $^{\mu}$ L of aqueous hum or in conical test tube, 2^{μ} L working standard solution of CsD and 1 mL ether were added successively. The tube was vortex-mixed for 60s and centrifuged for 2 m in at 3000 rpm. The supermatant layer was transferred to another clean tube and evaporated in water bath at 60°C. After reconstitution with 20 $^{\mu}$ L mobile phase, 10^{μ} L volume was injected into the chromatograph.

3 Results and conclusion

3. 1 Chrom a tog raphic behavior

The selectivity of the HPLC method was checked by analyzing different independent blank aqueous humor samples from rabbit eyes respectively. Representative chromatograms showed in Fig 1 indicate that no endogeneous interfering peaks at the retention time of CsA were found.

3. 2 Linearity

The linearity was evaluated with six blank aqueous humor spiked with various amounts of working standard solution containing 25, 50, 100, 200, 400, and 500 ng· mL⁻¹ of CsA. For each concentration three measurements were performed and

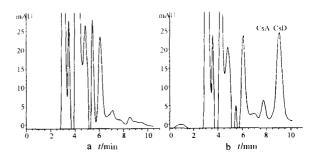


Fig 1 Chromatograms of blank aqueous humor (a) and controlled sample (b) of rabbit eyes

图 1 房水中环孢素 A的 HPLC图

a.空白房水色谱图 b.样品色谱图

calibration curves based on the peak height ratio of CsA and CsD versus nominal concentrations of CsA expressed in ng \cdot mL $^{-1}$ were constructed. The equation was $y=0.694391\,x$ with the correlation coefficient of 0.9986 and was calculated by least-square linear regression analysis packed in the chem station.

3.3 Accuracy and recovery

In the accuracy evaluation, three working standard solutions in aqueous hum or covering the calibration range and containing 25, 100 and 500 ng· mL¹ of CsA were prepared as described in the pretreatment section previously. Five consecutive measurements were performed for each concentration within the same day. Relative recovery was calculated by comparing the results from the standard curve above and the theoretical concentrations. Extraction recovery was calculated by comparing the heights of CsA from aqueous hum or spiked with working standard solution and those resulting from the direct sampling of working standard solutions prepared in mobile phase having the same theoretical concentrations of CsA considering the folding effect and complete recovery of the pretreatment. The individual and the overall mean percent recovery were calculated in Table 1 and a RSD value of maximum 15% was set as the acceptance criteria.

Tab 1 Recovery of the method for analysis of CsA in aqueous humor

表 1 眼前房水 CsA回收率 (n=5)

Concentration	Relative recovery		Absolute recovery	
(ng• mL-1)	recovery*	RSD (%)	recovery*	RSD (%)
25	104.4 ±10.1	9.7	86 ±7.2	8.4
100	99.4 ±6.1	6.1	88 ±6.8	7.7
500	98.46 ±4.36	4.4	83 ±7.3	8.8

Note: * calculated from peak height ratio (CsA: CsD) and expressed as recovery in percent, n=5, $\overline{x}\pm s$.

注: * 以峰高比计算 (CsA: CsD),以百分数表达, n = 5, $\overline{x} \pm s$.

3.4 Reproducibility

The within-day and day-to-day analytical reproducibility were determined by analyzing three different working standard solutions in aqueous humor having 25, 100 and 500 ng• mL⁻¹ of

CsA showed in Table 2. Five consecutive measurements were performed for each concentration within the same day and on five different days. The acceptance criteria for within-day and day-to-day R. S. D. values were not more than 15%.

3. 5 Sensitivity

The sensitivity in tems of detection and quantification limits were determined by using series of different working standard solutions in aqueous humor varying from 5 to 30 ng• mL⁻¹ of CsA, which were prepared as described in the standard solutions and pretreatment sections. The concentration measured with suitable accuracy and precision was accepted as the limit of quantification. The limit of detection at a signal-to-noise ratio of about 3:1 was regarded as the lowest detectable amount which was 10 ng• mL⁻¹.

Tab 2 Within-day and between-day reproducibility of the method for analysis of CsA in aqueous humor

表 2 精密度测定结果 (n=5)

Concentration	added-	W ith in-day		Be tween-day	
(ng• mL-1)		found*	RSD (%)	found* (%)	RSD (%)
25	25	26.1 ±2.52	9.7	26.7 ±3.56	13.5
100	100	100.2 ±6.93	6.9	96.8 ±9.97	10.3
500	500	506.5 ±21.6	4.3	504.7 ±29.1	5.8

Note: Calculated from peak height ratio (CsA: CsD) and expressed as found amount in percent, n = 5, $\bar{x} \pm s$.

注: "以峰高比计算(CsA: CsD),以百分数表达, n=5, $\overline{x}\pm s$

4 Discussion

Most of the HPLC methods [3-7] use extensive sample cleanup procedure following deprote inization that complicates HPLC assay of CsA in the whole blood. This may explain why most of the laboratories prefer to use immunoassay methods. In the present study, it is demonstrated that CsA in aqueous hum or could be analyzed by HPLC following only one-step extraction.

Commercially available HPLC columns show great differences in their chromatographic behavior. Columns with different packing materials such as C18, C8, and CN were tested and all were found suitable according to the refference [8]. Columns like CN should be operated at 60°C with the mobile phase of 50% acetonitrile, while 70°C and 75% acetonitrile should be used for C18 or C8 columns. To improve the sensitivity, analytical narrow-bore column of C18 controlled at 75°C was applied in this study with better performance. The detective wave length of 200 nm was used in this study which was different from reference but showed strongest absorption. The reduction of flow rate from usual 1 m L · m in · 1 to 0.1 m L · m in · 1 causes the probability of improving sensitivity greatly which makes it possible to measure CsA in aqueous hum or.

The storage condition is usually considered important when

validating a method. As is reported that freezing of blood samples increases interferences dramatically and should be avoided. Several studies show that CsA is stable in blood samples for $2 \sim 3$ months when stored at $1 \sim 4^{\circ}C^{\{8\}}$. In this study, even though stored up to more than 3 months at $4^{\circ}C$ or below $0^{\circ}C$, the number and intensity of interfering peaks were not increased and the CsA was still well separated in aqueous humor samples. Representative chromatograms obtained with CsA are shown in Fig. 1. The CsA peak, which had a retention time of 7.8 m in, was well resolved and free of interference from endogenous compounds in the aqueous humor. One fact should be noted that polypropylene tubes can not be used for pretreatment to avoid interfering additives which have much similar chromatographic behavior as CsA in them.

In conclusion, the HPLC method described here is simple, sensitive, reproducible and is applicable to pharmacokinetic studies of CsA in aqueous humor of rabbit eyes during topical the rapy.

References

- [1] Xie LX, Shi WY, Wang ZY, et al. Prolongation of comeal allograft survival using cyclosporine in a polylactide-co-glycolide polymer [J]. Comea, 2001, 20 (7): 748.
- [2] Theng JT, Ti SE, Zhou L. Pharmacokinetic and toxicity study of an intraocular cyclosporine DDS in the anterior segment of rabbit eyes[J]. Invest Ophthalmol Vis Sci, 2003. 44(11): 4895.
- [3] Jayasimha N. Murthy, Randall. Cyclosporine metabolite cross-reactivity in different cyclosporine assays[J]. Clin Biochem, 1998, 31(3):159.
- [4] Masri M, Rizk S, Andrysek T, et al. Cyclosporine blood level monitoring. Cross-reactivity of anti-cyclosporine A monoclonal with its sulphate metabolite: an in vitro study [J]. Mol Immunol, 2003, 39(17-18): 1059.
- [5] Dusei DJ, Hackett LP, Chswell GM, et al. Comparison of cyclosprine measurement in whole blood by high performance liquid chromatography, monoclonal flurescence polarization immunoassay and monoclonal enzyme multiplied immunoassay[J]. Ther Drug Moit, 1992, 14 (4): 327.
- [6] Rustum AM. Estimation of cyclosporin in whole blood by simple and rapid reversed-phase HPLC utilizing a salting out extraction procedure[J]. J Chromatogr Sci, 1990, 28(11): 594.
- [7] Zaghloul AA, Hussain A, Khan MA, et al. Development of a HPLC method for the determination of cyclosporin-A in rat blood and plasma using naproxen as an internal standard[J]. J Pharm Biomed Anal, 2003, 31 (6):1101.
- [8] Hosse in Am ini, Abolhassan Ahmadiani. Simple determination of cyclosporine in human whole blood by high-performance liquid chromatography J. J. J. Chromatogr. B, 2003, (795): 209.

收稿日期:2004-03-11